Scrutinizing of Trace Elements and Antioxidant Enzymes Changes in Barki Ewes Fed Salt-Tolerant Plants under South Sinai Conditions

Hanan, Z. Amer; Ibrahim, N. H.; Donia, G. R.; Younis, F. E. and Shaker, Y. M.

1Animal and Poultry Physiology Department, Animal and Poultry Division, Desert Research Center, El-Matarya St., Egypt
2Animal and Poultry Health Department, Animal and Poultry Division, Desert Research Center, El-Matarya St., Egypt
3Department of Biology- Faculty of Science- Jazan University- Kingdom of Saudi Arabia
nagy_drc7777@yahoo.com

Abstract: In attempt to monitor the pattern of trace elements and antioxidant changes in ewes as a result of feeding silage of salt tolerant plants during different physiological status under South Sinai conditions. Forty two Barki ewes were randomly divided into two equal groups (21 each). The first group (control, G1) was fed berseem hay while the second group (G2) was fed silage form of salt tolerant plants and concentrates feed mixtures. This experimental was carried out at South Sinai Station (Ras Sudr) belonging to Desert Research Center (DRC), Ministry of Agricultural and Land Reclamation, Egypt.

Blood samples were collected from the all animals during dry, gestation (early, mid, late) and early lactation periods. The profiles of trace elements (Cu, Se, Mn and Zn), malondialdehyde (MDA), lipid peroxidation and oxidative stress markers [total antioxidant capacity (TAC), antioxidant catalase (CAT) enzymes] were analyzed in plasma. On the other hand, antioxidant enzyme (SOD and GPX) activities were analyzed in erythrocyte.

The obtained results declared that there were significant differences in plasma levels of trace elements (Cu, Mn, Se and Zn) and consequently antioxidant enzymes (SOD and GPX) between treatment groups. On the other hand, pregnancy and lactation constituted the most oxidative stress facing the animals of the two groups since oxidative stress index (MDA) was increased and TAC was decreased and were significantly different in treatments and physiological status.

It could be concluded that feeding silage form of salt tolerant plants was not harmful for desert Barki ewes raised under semi-arid condition of South Sinai. Furthermore, pregnancy and lactation periods constituted oxidative stresses on animals even fed traditional or untraditional (salt tolerant plants) diets. So, it is recommended that supplementing trace elements diet in order to improve antioxidant status (defense system) which consequently enhances growth performance and animal productivity.


Keywords: Salt tolerant plants, Barki ewes, pregnancy, trace elements, lipid peroxidation, oxidative stress, antioxidant enzymes,
peroxidation, protein denaturing and DNA mutation. Evidences suggest that membranes are the primary sites of oxidative stress because ROS can react with unsaturated fatty acids causing peroxidation of essential membrane lipids in plasma membrane or intracellular organelles (Esfandiari et al., 2007). Polyunsaturated fatty acids present in membrane phospholipids are the main target substrates for oxygen radical activity which results in disorganization of cell framework and function (Nazifi et al., 2010a). Peroxidation of plasma membrane leads to the leakage of cellular contents, rapid desiccation and cell death. Lipid peroxides derived from polyunsaturated fatty acids are unstable and are decomposed to form a series of compounds, including malondialdehyde (MDA). The quantization of MDA is widely used as an indicator of lipid peroxidation and oxidative stress in cells and tissues (Simek et al., 2006). The cells have evolved a number of counteracting antioxidant defenses. Antioxidant enzymatic activities are of utmost importance because they may indicate how tissue or organ might respond to oxidative stress in oxidizing environment. The natural defense mechanisms against free radicals consist of enzymatic antioxidants like glutathione peroxidase (GSH-Px), superoxide dismutase (SOD), catalase (CAT) and non-enzymatic antioxidants like glutathione (GSH), ascorbate, urate, vitamin E and beta carotene (Erisir et al., 2009). SOD is the first line of defense against ROS and is active in catalyzing detoxification of superoxide radical. The hydrogen peroxide generated in this reaction is restored to water in the presence of CAT and GPX (Nazifi et al., 2010a).

Therefore, this study was carried out so as to monitor the changes in some trace elements (Cu, Zn, Mn and Se), malondialdehyde (MDA), lipid peroxidation and oxidative stress markers (total antioxidant capacity (TAC), antioxidant catalase (CAT) enzymes) and antioxidant enzyme (SOD and GPX) activities which may occur during different physiological stages in Barki ewes fed salt tolerant plants raised under semi-arid conditions of South Sinai, Egypt.

2. Materials and Methods

This investigation was undertaken at South Sinai Station (Ras Sudr) which belongs to Desert Researcher Center, Ministry of Agriculture and Land Reclamation, Egypt, in order to designate the effect of feeding salt tolerant plants on changes in antioxidant enzymes and its relation with plasma profiles of trace elements (Cu, Zn, Mn and Se) during different physiological status.

Forty two Barki ewes aging 2.5-3 years old and averaging 41.50± 4.85 kg body weight were divided into two equal groups. The first group was fed berseen hay (Trifolium alexandrinum, 4th cut) and served as control while the second one was fed silage form of salt tolerant plants (Atriplex halimus, 50%; fodder beet, 25%; Pearl millet, 15% and Carthamus tinctorius hay, 10%). Experimental animal were fed their nutrient requirements according to Kearl (1982).

The trace elements in terms of copper (Cu), manganese (Mn), selenium (Se) and zinc (Zn) were analyzed in the both rations of the two groups using microwave digestion technique (1.5 ml of sample + 8 ml of HNO3 65%, 2 ml of H2O2 30% in a closed Teflon vessel under high temperature and pressure control) as reported by Littlejohn et al., (1991). The metals were determined by spectroscopic methods, Flame photometer and Inductively Coupled Plasma Mass Spectroscopy (ICP-MS) in the central Lab of Desert Research Center (Water and Soil Analysis Unit).

<table>
<thead>
<tr>
<th>Table (1): Trace elements concentration (ppm) in the experimental rations</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control group: Copper</td>
</tr>
<tr>
<td>------------------------</td>
</tr>
<tr>
<td>9.40</td>
</tr>
<tr>
<td>Silage group: Copper</td>
</tr>
<tr>
<td>13.26</td>
</tr>
</tbody>
</table>

Blood samples were collected into heparinized tube (10 ml) from all experimental animals. Heparinized blood was centrifuged at 3000 rpm for 15 min. at 4°C. Pipette off plasma without disturbing the white buffy layer. Plasma was kept and stored at -80°C pending analysis of trace elements, lipid peroxide (MDA), total antioxidant capacity (TAC) and catalase (CAT) enzymes.

Trace elements (Cu, Zn, Mn and Se) levels analysis in plasma were determined by flame atomic absorption spectrophotometer (Pye-Unicam SP9). Lipid peroxidation was assayed by measuring the level of malondialdehyde (MDA) by the method of Ruiz-Larrea et al. (1994) using thiobarbituric acid (TBA). The acid reacts with MDA to form a stable pink color with maximum absorption at 532 nm and expressed as nmol/ml.

TAC (mU/l) and catalase (U/l) were measured by colorimetric techniques using a commercially available kit (Bio-diagnostic, Egypt) according to the method of Koracevic et al. (2001) and Aebi (1984), respectively.
For determination of superoxide dismutase enzyme (SOD), 0.5 ml of heparinized whole blood was centrifuged for 10 min. at 4000 rpm at 4°C and then plasma aspirated off. Then red blood cells washed four times with 3 ml of 0.9% saline solution (NaCl), centrifuged for 10 min. at 4000 rpm after each wash. The washed centrifuged erythrocytes should then be made up to 2.0 ml with cold redistilled water. Mixed and left to stand at +4°C for 15 min. and stored at -80°C until analysis. The lysate is diluted with distilled water (50 fold), so the % inhibition falls between 30% and 60%. SOD measured calorimetrically using a commercially available kit (Bio-diagnostic., Egypt) according to the method of Nishikimi et al., (1972) and expressed as U/L.

For determination of glutathione peroxidase enzyme (GPX), the red blood cells collected by centrifugation of heparinized whole blood (4000 rpm x 10 min. at 4°C) then plasma drawn off. Erythrocytes washed once (one time) with 10 volumes of cold saline. The red blood cell pellets lysed by adding 4 volumes of cold deionized water to the estimated pellet volume, then the red cell stroma was removed by centrifuging (4000 rpm x 10 min. at 4°C). The resulting clarified supernatant was collected and stored at -80°C until assay. GPX measured calorimetrically using a commercially available kit (Bio-diagnostic., Egypt) according to the method of Paglia and Valentine (1967) and expressed as mU/mL.

Experimental data were analyzed using General Linear Model Procedure (SAS, 2004).

3. Results and Discussion

The obtained results demonstrated that feeding salt tolerant plants silage lowered (P< 0.01) the mean values of plasma and manganese than control group (Table 2) although salt tolerant plants silage contained higher levels of manganese (Mn) (Table 1). Unfortunately, concentrations of Mn in plasma are not good indicators of Mn intake. Concentrations of Mn in the red blood cells are higher than in plasma and have been used to assess status (Hidiroglou et al., 1978). Dietary Mn affects the concentration of Mn in bones and other tissues (Kincaid, 1999). Exhibiting the same trend, selenium (Se) levels in the blood serum was lower (P<0.01) in ewes fed salt tolerant plants silage than in their counterparts fed the traditional ration (Table 2). These results might be due to the low content of (Se) in salt tolerant plants silage ration. Concentrations of Se in whole blood are responsive to Se intake (Levander, 1986).

Contrariwise, animals salt tolerant silage group achieved higher (P< 0.01) copper (Cu) and zinc (Zn) mean values than control group (Table 2). The higher values of copper (Cu) in salt tolerant plants silage group could be attributed to the higher Cu intake in the diet (Table 1). Ashton (1970) reported that copper levels in tissues and body fluids depend on diet, state of health, age and sex. Copper is a mineral element that activates several enzyme systems, and though in less numbers than Zn, it is considered an essential nutrient (Minatel and Carfagnini, 2000). The physiological role of Cu in the organism is related to several functions, which include cellular respiration, bone formation, connective tissue development, and essential catalytic cofactor of some metallo-enzymes, among other (McDowell, 2003 and Underwood and Suttle, 2003).

Unexpectedly, zinc concentrations in plasma of salt tolerant plants animals exceeded (P< 0.01) their counterparts of control group (Table 2) although the low levels of Zn intake (Table 1). This might be that animals fed salt tolerant plants were more efficient to utilize the low Zn intake. Elnageeb and Abdelatif (2010) suggested that a combination of low nutritional status and pregnancy in non-supplemented ewes may increase the efficiency of utilization of ingested Zn. The major part of the total body Zn is in the bones and competes with Cu for absorption from the intestinal tract (Kargin et al., 2004). The need for Zn in most animals is based on its influence on enzymes and proteins and their activities, that are linked to vitamin A synthesis, carbon dioxide (CO2) transport, collagen fiber degradation, free radical destruction, membrane stability of red blood cells, metabolism of essential fatty acids, carbohydrate metabolism, protein synthesis and metabolism of nucleic acids, among others (Powell, 2000; McCall et al., 2000; Stefanidou et al., 2006 and Rubio et al., 2007).

Concerning the effect of physiological status, the present results showed that pregnancy and lactation influenced the levels of serum micro-minerals of ewes fed silage of salt tolerant plants (Table 2). Pregnancy and lactation constituted metabolic stress, associated with alterations in the minerals profile dependent on the reproductive status of small ruminants. Metabolism of mineral elements plays a significant role in the regulation of physiological functions of pregnancy and lactation. Moreover, substantial losses of body minerals occur during pregnancy and lactation (Ceylan et al., 2009; Elnageeb and Adelatif, 2010). The concentration of minerals varies in blood as a result of interactions between those nutrients, transfer of nutrients to the fetus and initiation of milk synthesis (Kincaid, 2008). Pregnancy presents a considerable stress to trace mineral homeostasis in mammals (Mills and Davies 1979).
Table (2): Means of some trace elements concentrations of the experimental groups as affected by feeding silage of salt tolerant plants during different physiological status under South Sinai conditions

<table>
<thead>
<tr>
<th>Minerals</th>
<th>Treat</th>
<th>Dry</th>
<th>Early</th>
<th>Mid</th>
<th>Late</th>
<th>Lactation</th>
<th>Overall</th>
<th>±SE</th>
<th>T</th>
<th>S</th>
<th>TxS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cu, ppm</td>
<td>T1</td>
<td>1.96</td>
<td>1.54</td>
<td>1.07</td>
<td>0.94</td>
<td>1.67</td>
<td>1.44</td>
<td>0.07</td>
<td>0.11</td>
<td>0.15</td>
<td></td>
</tr>
<tr>
<td></td>
<td>T2</td>
<td>2.00</td>
<td>2.36</td>
<td>2.36</td>
<td>2.10</td>
<td>0.94</td>
<td>1.95</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Overall</td>
<td>1.98</td>
<td>1.95</td>
<td>1.71</td>
<td>1.52</td>
<td>1.30</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mn, ppm</td>
<td>T1</td>
<td>0.137</td>
<td>0.499</td>
<td>0.855</td>
<td>0.675</td>
<td>0.163</td>
<td>0.47</td>
<td>0.02</td>
<td>0.03</td>
<td>0.04</td>
<td></td>
</tr>
<tr>
<td></td>
<td>T2</td>
<td>0.487</td>
<td>0.481</td>
<td>0.233</td>
<td>0.199</td>
<td>0.154</td>
<td>0.31</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Overall</td>
<td>0.319</td>
<td>0.490</td>
<td>0.543</td>
<td>0.437</td>
<td>0.158</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Zn, ppm</td>
<td>T1</td>
<td>7.05</td>
<td>6.73</td>
<td>10.22</td>
<td>8.25</td>
<td>7.20</td>
<td>7.89</td>
<td>0.29</td>
<td>0.46</td>
<td>0.65</td>
<td></td>
</tr>
<tr>
<td></td>
<td>T2</td>
<td>6.34</td>
<td>6.38</td>
<td>12.59</td>
<td>11.22</td>
<td>8.57</td>
<td>9.02</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Overall</td>
<td>6.69</td>
<td>6.56</td>
<td>11.40</td>
<td>9.73</td>
<td>7.88</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Se, ppm</td>
<td>T1</td>
<td>0.035</td>
<td>0.076</td>
<td>0.116</td>
<td>0.101</td>
<td>0.225</td>
<td>0.11</td>
<td>0.01</td>
<td>0.02</td>
<td>0.02</td>
<td></td>
</tr>
<tr>
<td></td>
<td>T2</td>
<td>0.104</td>
<td>0.075</td>
<td>0.052</td>
<td>0.055</td>
<td>0.033</td>
<td>0.063</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Overall</td>
<td>0.069</td>
<td>0.075</td>
<td>0.084</td>
<td>0.078</td>
<td>0.129</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

T1: animals fed berseem hay  
T2: animals fed salt tolerant plants silage

In the same column, means in a certain item having the same small letter do not differ significantly.
In the same row, means in a certain item having the same capital letter do not differ significantly.

Serum Cu level was lower during lactation compared to the respective values obtained pre and during pregnancy (Table 2). This could be related to the response of the ewes to the needs of the foetus by increasing mobilization of stored Cu for the development of the nervous system (Elnageeb and Abdelatif, 2010).

The present results demonstrated that the differences in Zn levels among the periods of dry, pregnancy and lactation were significant (P< 0.01). However, the levels of Zn were higher in mid, late pregnancy and lactation period as compared to the dry period (Table 2). These results could be attributed to the increase in the rate of accumulation of Zn in the foetus. Williams et al. (1972) reported that the developing foetus accumulates 1 to 2 mg of Zn/ day and the pregnant ewe increases the demands for Zn towards the end of pregnancy. However, Elnageeb and Abdelatif (2010) reported that no significant changes were observed in Zn level during the experimental periods (dry, pregnancy and lactation periods). The serum Zn level was slightly higher during pregnancy compared to the value obtained during early flushing period. There is also evidence in sheep and cattle that the Zn status and intake affect Zn absorption from the gut (Kirchgeissner, 1976 and Suttle, 1988).

Concerning the effect of physiological status on the manganese level, the obtained results demonstrated that pregnancy increased (P<0.01) the Mn level from the dry period to reached its peak at mid- pregnancy then it decreased gradually till the lactation period which had the lowest Mn values (Table 2). Similarly, El-Tohamy et al. (1986) reported lower plasma manganese in non-pregnant camels. However, according to the authors, no variation owing to pregnancy was observed, contrary to other trace elements.

During pregnancy and lactation, the concentration of Se decreased in ewes fed silage of salt tolerant plants as compared with control ewes that fed berseem hay (Table 2) which might be attributed to the low Se concentration in salt tolerant plants (Table 1). This may be explained by the concentration of selenium in plants varies widely and depends on the selenium content and characteristics of the soil (Pechová et al., 2008). The selenium concentration in soil is low in many parts of the world including South Sinai Research Station where it is poorly available and incapable of providing the required amount to animals because of the presence of the antagonistic relationship between water irrigation salinity and Se available in the soil (Sadek, 1995 and Fahmy et al., 2009).

The detection of free radical damage and the protection against it has become very important in the studies related to ruminant production and reproduction as the level of lipid peroxidation and antioxidant status give complementary information about the metabolic status of the animal rather than metabolic parameters alone (Castillo et al., 2003).

The obtained results, as shown in Table (3), revealed that malondialdehyde (MDA) level was found to be significantly (P< 0.01) increased in the
two experimental groups along the advanced of pregnancy. This gradual increase with the progression of pregnancy was associated with decreases antioxidant enzyme levels; total antioxidant capacity (TAC), antioxidant catalase (CAT) enzyme, superoxide dismutase (SOD) and glutathione peroxidase (GPX). The maximum level of lipid peroxidation was observed in late pregnancy and early lactation in control ewes that fed berseem hay comparing with the level of MDA in ewes fed silage of salt tolerant plants in late pregnancy. This finding is in agreement with the findings of Toescu et al., (2002), Upadhyaya et al., (2005) and Patil et al., (2006 and 2007) who reported that markers of lipid peroxidation (MDA) to be increased during the progression of normal pregnancy. MDA is considered the final product of lipid peroxidation and a marker of oxidative stress. In the same time, trace elements showed the same trend of decrease in ewes fed silage of salt tolerant plants especially Se in all experimental periods, Cu in early lactation and Mn in late pregnancy and early lactation (Table 2). It is known that various kinds of stress such as salinity, pregnancy and lactation accelerate the production of reactive oxygen species and oxidative stress (Górecka et al., 2002). These species of oxygen are highly cytotoxic and can seriously react with vital biomolecules such as lipids, proteins, nucleic acid, etc., causing lipid peroxidation, protein denaturing and DNA mutation (Esfandiar et al., 2007). In health, reactive oxygen species (ROS) and antioxidants remain in balance but this balance is disrupted in cases of oxidative stress (Aurousseau et al., 2006).

The rise of oxidative stress markers could be due to pregnancy and early lactation which are considered as stressful stages accompanied by a high metabolic demand and elevates the requirements for tissue oxygen (Patil et al., 2007 and Idonije et al., 2011) and causes an increase of reactive oxygen species production. This could be explained by the fact that 80% of foetus growth occurs in the last 2 months of pregnancy, so ewes exhibit a dramatic increase in metabolism during this period (Cristian and Jauhianinan, 2001). The rapid growth of foetus during the last 6 weeks of pregnancy, causes an increase in fatty acid consumption from the mother’s fat reserve and production of hydrogen peroxide that has been enhanced by intense lipolysis and mobilization of fatty acids from the body deposits during pregnancy (Oztabay et al., 2005 and Rezapour and Roudbaneh, 2011) and during lactation in order to sustain the lactogenesis (Adela et al., 2006). Moreover, the placenta is a major source of oxidative stress because of its enrichment with polyunsaturated fatty acids (PUFA) (Gitto et al., 2002).

In addition to pregnancy, salinity of salt tolerant plants is another stressor on ewes. The soils where halophytes normally grow becomes more saline due to rapid evaporation of water particularly during

| Table (3): Means of malondialdehyde and antioxidant enzymes concentrations of the experimental groups as affected by feeding silage of salt tolerant plants during different physiological status under South Sinai conditions |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Minerals        | Treat           | Dry  | Early | Mid  | Late | Lactation | Overall | ±SE  | T   | S   | TxsS |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| MDA (nmol/ml)   | T1              | 0.78<sup>ab</sup> | 1.26<sup>ab</sup> | 1.30<sup>ab</sup> | 1.33<sup>ab</sup> | 1.33<sup>ab</sup> | 1.20<sup>a</sup> | 0.02** 0.02** | 0.03<sup>NS</sup> |
|                 | T2              | 0.68<sup>ab</sup> | 1.14<sup>ab</sup> | 1.12<sup>ab</sup> | 1.23<sup>ab</sup> | 1.13<sup>bh</sup> | 1.06<sup>b</sup> |                 |                 |                 |
|                 | Overall         | 0.75<sup>c</sup> | 1.20<sup>b</sup> | 1.21<sup>ab</sup> | 1.28<sup>a</sup> | 1.23<sup>ab</sup> |                 |                 |                 |                 |
| TAC (mU/L)      | T1              | 1.32<sup>ab</sup> | 1.06<sup>bc</sup> | 0.70<sup>ab</sup> | 1.04<sup>ab</sup> | 1.14<sup>ab</sup> | 1.13<sup>a</sup> | 0.02** 0.02** | 0.03<sup>*</sup> |
|                 | T2              | 1.35<sup>ab</sup> | 1.02<sup>ab</sup> | 0.83<sup>bc</sup> | 0.95<sup>ab</sup> | 1.04<sup>ab</sup> | 1.04<sup>b</sup> |                 |                 |                 |
|                 | Overall         | 1.33<sup>c</sup> | 1.04<sup>bc</sup> | 0.95<sup>d</sup> | 1.00<sup>cd</sup> | 1.09<sup>b</sup> |                 |                 |                 |                 |
| Catalase (U/L)  | T1              | 203.02         | 153.20          | 148.53          | 70.21           | 61.80          | 127.35          | 2.30<sup>NS</sup> | 3.64<sup>**</sup> | 5.14<sup>NS</sup> |
|                 | T2              | 204.46         | 170.65          | 139.05          | 78.04           | 57.14          | 129.87          |                 |                 |                 |
|                 | Overall         | 203.74<sup>d</sup> | 161.93<sup>b</sup> | 143.79<sup>c</sup> | 74.12<sup>d</sup> | 59.47<sup>e</sup> |                 |                 |                 |                 |
| SOD (U/L)       | T1              | 235.12<sup>ab</sup> | 244.34<sup>ab</sup> | 222.78<sup>ab</sup> | 197.40<sup>ab</sup> | 187.58<sup>ab</sup> | 217.44<sup>a</sup> | 3.16** 4.99** | 7.06<sup>**</sup> |
|                 | T2              | 246.36<sup>ab</sup> | 210.36<sup>ab</sup> | 171.25<sup>bc</sup> | 189.59<sup>ab</sup> | 215.92<sup>ab</sup> | 206.70<sup>b</sup> |                 |                 |                 |
|                 | Overall         | 240.74<sup>d</sup> | 227.35<sup>c</sup> | 197.02<sup>ab</sup> | 193.49<sup>ab</sup> | 201.75<sup>b</sup> |                 |                 |                 |                 |
| CPX (mU/L)      | T1              | 453.91<sup>ab</sup> | 402.04<sup>ab</sup> | 252.89<sup>bc</sup> | 376.10<sup>ab</sup> | 233.44<sup>ac</sup> | 343.67<sup>a</sup> | 5.62** 8.88** | 12.56<sup>**</sup> |
|                 | T2              | 434.46<sup>ab</sup> | 337.19<sup>ab</sup> | 149.14<sup>b</sup> | 226.96<sup>bd</sup> | 207.50<sup>ad</sup> | 271.05<sup>b</sup> |                 |                 |                 |
|                 | Overall         | 444.19<sup>d</sup> | 369.61<sup>b</sup> | 201.02<sup>d</sup> | 301.53<sup>c</sup> | 220.47<sup>cd</sup> |                 |                 |                 |                 |

T1: animals fed berseem hay  
T2: animals fed salt tolerant plants silage

In the same column, means in a certain item having the same capital letter do not differ significantly.
In the same row, means in a certain item having the same small letter do not differ significantly.
summer, therefore, surface of the soil tend to have higher soil salinity and higher water potential (Khan and Gul, 2002). The adverse effects of the salt on cell membranes are results of the accumulating toxic ions and ROS (Cicerali, 2004) and evidence suggests that membranes are the primary sites of salinity injury to cells and organelles because ROS can react with PUFA and results in disorganization of cell framework and function and cause peroxidation of essential membrane lipids in plasma membrane or intracellular organelles. Peroxidation of plasma membrane leads to the leakage of cellular contents, rapid desiccation and cell death. Lipid peroxidation is an indicator of oxidative stress in cells and tissues (Esfandiari et al., 2007).

The result of the present study indicated that there was a negative relationship between antioxidant enzyme activities and lipid peroxidation or MDA content (Table 3). Enzymes with important antioxidant functions include: i) superoxide dismutase (SOD), which catalyses the dismutation of superoxide radical to hydrogen peroxide and water, ii) catalase (CAT), which catalyses the breakdown of hydrogen peroxide to oxygen and water, and iii) glutathione peroxidase (GPX), which facilitates the destruction of both hydrogen peroxide and organic peroxides. SOD is the first line of defense against ROS and is active in catalyzing detoxification of superoxide radical (Nazifi et al., 2010a). The hydrogen peroxide generated in this reaction is restored to water in the presence of CAT and GPX.

Several studies have indicated that antioxidative defense system is modified during normal pregnancy. Late-pregnant ruminants tend to have raised lipoperoxidative processes as a consequence of increased production of free oxygen radicals and therefore have a low antioxidative status (Dimri et al. 2010). The susceptibility of cells to oxidative stress is a function of the overall balance between the degree of oxidative stress and the antioxidant defense capability. Decreased concentrations of catalase, SOD and GPX activities may reflect oxidative stress in pregnant ewes (Erisir et al., 2009).

It is worthwhile to mention that the decrease of antioxidant enzymes level during late pregnancy and early lactation in our study is dependent on trace elements profile where trace elements showed the same trend of decrease in both experimental groups; Se in all experimental periods, Cu in early lactation and Mn in late pregnancy and early lactation (Table 2 and 3).

Animals fed plants grown in selenium-deficient areas and not supplemented with minerals are vulnerable to oxidant stress (Steen et al., 2008). Many authors confirmed a positive correlation between GSH-Px activity and selenium concentration in whole blood where about 11.8% of total Se in the organism is bound in GSH-Px. (Awadeh et al., 1998) and Trávníček et al., (2008) proved a high correlation between Se content in the whole blood and GSH-Px activity in the blood of ewes. Our findings demonstrates that the correlation between GSH-Px activity and selenium concentration in the whole blood of ewes is very close and that GSH-Px activity could be considered as a good indicator and diagnostic tool for the determination of selenium status in sheep (Pavlata et al., 2012) and consequently in the evaluation of antioxidant status (Adela et al., 2006).

According to Sattar et al. (2007), pregnant animals are more susceptible to selenium deficiency than non-pregnant animals and maternal selenium concentrations and glutathione peroxidase activity fall during pregnancy (Mistry and Williams, 2011). This decrease in selenium status which is progressive as gestation proceeds may be partly attributed to hemodilution from the blood volume increase associated with pregnancy (Boskabadi et al., 2010). Moeini et al., (2011) reported that Se concentration decreased during the final 60 days of gestation, emphasizing the importance of se supplementation during late gestation

Humphries et al. (1983) revealed that in experimental copper deficiency in calf, plasma concentration of copper and SOD activity of erythrocytes severely decreased. Similarly, Nazifi et al. (2010b) found a positive correlation between plasma concentration of copper and SOD activity of erythrocytes because copper, zinc and magnesium are the main components of SOD that plays a vital role as an antioxidant and protects from oxidative stress (Nazifi et al., 2010b).

From the above results, it could be concluded that feeding desert animals; Barki ewes, silage form of salt tolerant plants was not hazardous under semiarid conditions of South Sinai. Furthermore, the pregnancy and lactation periods constitute oxidative stresses on animals even fed traditional or untraditional (salt tolerant plants) diets. So, we recommended supplementing trace elements to the diets so as to improve the antioxidants capacity (defense system) which consequently enhance growth performance and animal productivity.

**Corresponding author**

Ibrahim, N. H.

Animal and Poultry Physiology Department, Animal and Poultry Division, Desert Research Center, El-Mataryia St., Egypt

nagy_drc7777@yahoo.com

**Acknowledgment**

The authors would thank Prof. Dr. El-Shaeer, the coordinator and Prof. Dr. Badawy, the key person of
the regional project titled “Adaptation to climate changes in WANA marginal environments through sustainable crop and livestock diversification” which is funded by International Center for Biosaline Agricultural (ICBA), UAE for their financial support to achieve this work.

References


